

## LETTERS

## Assembly reflects evolution of protein complexes

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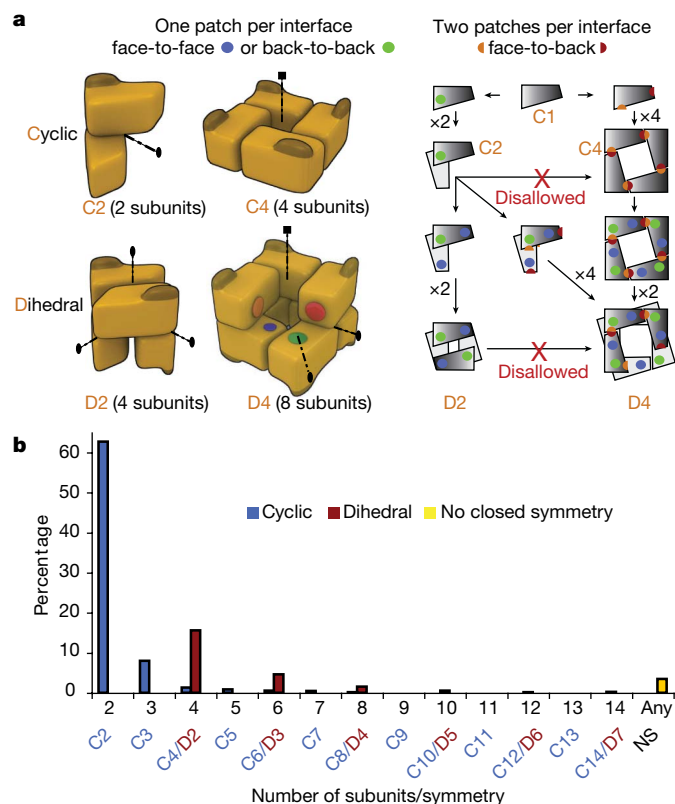
A homomer is formed by self-interacting copies of a protein unit. This is functionally important<sup>1,2</sup>, as in allostery<sup>3–5</sup>, and structurally crucial because mis-assembly of homomers is implicated in disease<sup>6,7</sup>. Homomers are widespread, with 50–70% of proteins with a known quaternary state assembling into such structures<sup>8,9</sup>. Despite their prevalence, their role in the evolution of cellular machinery<sup>10,11</sup> and the potential for their use in the design of new molecular machines<sup>12,13</sup>, little is known about the mechanisms that drive formation of homomers at the level of evolution and assembly in the cell<sup>14</sup>. Here we present an analysis of over 5,000 unique atomic structures and show that the quaternary structure of homomers is conserved in over 70% of protein pairs sharing as little as 30% sequence identity. Where quaternary structure is not conserved among the members of a protein family, a detailed investigation revealed well-defined evolutionary pathways by which proteins transit between different quaternary structure types. Furthermore, we show by perturbing subunit interfaces within complexes and by mass spectrometry analysis<sup>15</sup>, that the (dis)assembly pathway mimics the evolutionary pathway. These data represent a molecular analogy to Haeckel's evolutionary paradigm of embryonic development, where an intermediate in the assembly of a complex represents a form that appeared in its own evolutionary history. Our model of self-assembly allows reliable prediction of evolution and assembly of a complex solely from its crystal structure.

Although homomers are central to biology, only anecdotal knowledge exists on their principles of evolution and assembly, and no unifying theory has been proposed. Large increases in structural data in recent years, however, have enabled us to study quaternary structure or spatial arrangement of subunits on a data set of 5,375 unique structures. This data set is ~tenfold greater than any studied previously<sup>16</sup> (Methods). On the basis of this data set, we quantify how often proteins change their quaternary structure, and identify the evolutionary routes taken to do so. Subsequently, as evolution of a complex can be viewed as assembly over a long timescale, we compare evolutionary routes with (dis)assembly routes probed by mass spectrometry.

Homomers can be separated into two main classes of open or closed symmetry. The first class corresponds to open structures that would polymerize to infinity in the absence of limiting factors. Such assemblies (for example, tubulin and actin) are rare in our data set (3%), probably because their innate dynamic character renders them difficult to crystallize. In contrast, closed symmetries are finite in space, and most homomers adopt either cyclic or dihedral symmetry (Fig. 1a), with only a small fraction (1%) having cubic symmetry (not shown). Throughout we denote  $C_n$  as a cyclic complex containing  $n$  subunits, and  $D_n$  as a dihedral complex containing  $2n$  subunits.

It has long been observed that smaller complexes are more abundant than larger ones, and even numbers of subunits are favoured over odd numbers<sup>8,9,17</sup>. Here we confirm this observation, with 62% of complexes being dimers. We quantify the different types of symmetries found in homomers and show that the abundance of complexes

with even numbers of subunits is due to the prevalence of dihedral complexes. Whenever an option exists for cyclic or dihedral, on average we find an 11-fold preference for dihedral complexes (Fig. 1b). There is an evolutionary explanation for this preference, as the probability that a dihedral complex evolved by random mutation should be higher than the probability for a cyclic complex for at least two reasons: first, at the level of individual interfaces, in dihedral complexes most interfaces are face-to-face (or back-to-back), whereas all interfaces in cyclics are face-to-back (Fig. 1a) and these are less likely to form by random mutation<sup>5,18</sup>; and second, at the level of whole complexes, evolution of dihedral complexes can take place

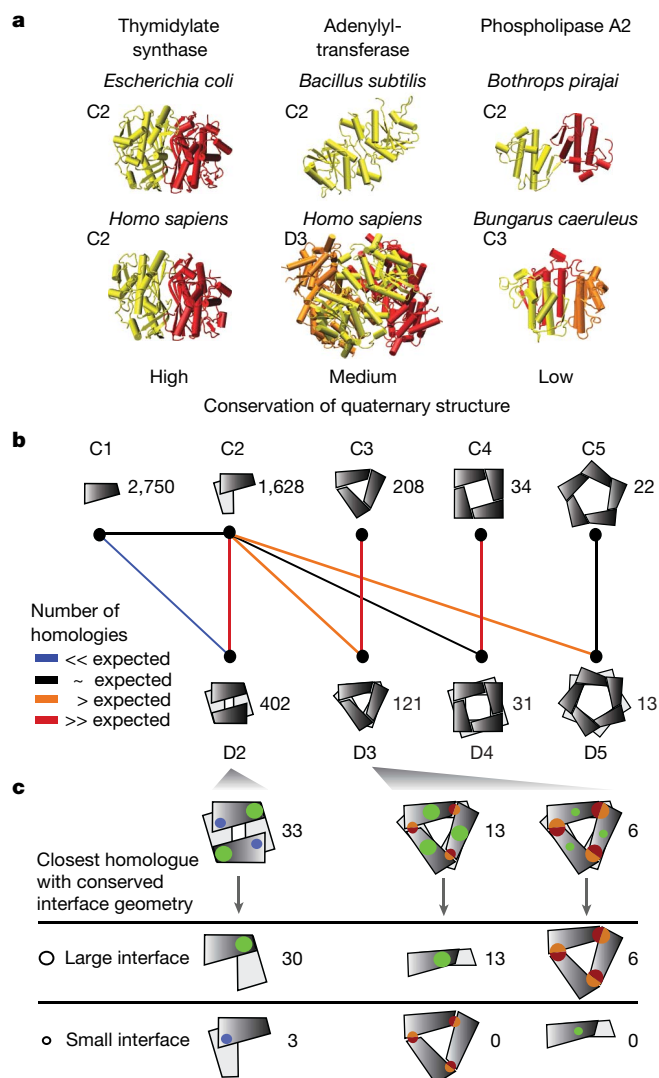


**Figure 1 | Abundance and properties of cyclic and dihedral symmetries.** **a**,  $n$  subunits in a cyclic complex are related by a single  $n$ -fold symmetry axis (dotted lines); ellipses and squares represent two- and four-fold axes, respectively. For a monomer to evolve towards a cyclic tetramer ( $C_4$ ), two complementary surfaces have to evolve simultaneously (red and orange patches). For a dihedral tetramer ( $D_2$ ), two different and self-complementary surfaces (green and blue patches) can evolve serially with an intermediate dimer ( $C_2$ ). **b**, The abundance of homomers with cyclic, dihedral, or no symmetry (3.5%). 62.7% are cyclic dimers ( $C_2$ ), 8% are cyclic trimers ( $C_3$ ), and 3.2% have higher order cyclic symmetry (from  $C_4$  to  $C_{14}$ ). Dihedral complexes dominate (22.6%) among complexes with  $\geq 4$  subunits.

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in multiple steps ( $C1 \rightarrow C2 \rightarrow D2$ ) whereas cyclics must evolve in one step ( $C1 \rightarrow C4$ , Fig. 1a).

Notably, dihedral and cyclic symmetries are geometrically related: a complex with  $D_n$  symmetry can be formed from  $n$  dimers with  $C2$  symmetry or from two  $n$ -mers with  $C_n$  symmetry<sup>19</sup> (Fig. 1a). If a protein complex has a particular symmetry, we find that homologues are likely to have the same symmetry type. More specifically, for sequence identities  $>90\%$ , conservation is nearly 100%, whereas in the range of 30–40% sequence identities, conservation is  $\sim 70\%$  (Supplementary Fig. 1). Proteins with different degrees of quaternary structure conservation are illustrated in Fig. 2a. Thymidylate synthase always exists as a dimer, adenylyltransferase is a dimer in *Bacillus subtilis* and a hexamer in human (trimer of *B. subtilis* dimers), whereas two phospholipase A2s have geometrically very distant quaternary structures.



**Figure 2 | Routes for homomer evolution.** **a**, Illustrative examples of different levels of quaternary structure conservation (termed high, medium and low). Thymidylate synthases (PDB accessions 1ajm and 1hw3) are both dimers. Adenylyltransferases (PDB accessions 1kam and 1kr2) differ in their number of subunits, with similar dimers (yellow) common to both quaternary structures. The dimer and trimer of venom-toxin phospholipase A2s are not related geometrically and the quaternary structures are therefore less conserved. **b**, Schematic illustration of large-scale analysis of quaternary structures; the number of unique complexes is indicated. Coloured lines indicate the significance in over-representation of shared homologous complexes (Methods). **c**, In 49 out of 52 cases, the largest interface is present in the dimeric or trimeric homologue, illustrated by size of interface patches.

When quaternary structure is not conserved, we speculate that pathways linking geometrically related symmetries represent both evolutionary and assembly routes. For example, a dihedral tetramer ( $D2$ ) can be described as a dimer of dimers, where a back-to-back dimerization patch forms a first dimer, and a second face-to-face dimerization patch forms the dimer of dimers. This is not true of a cyclic tetramer ( $C4$ ), where subunits interact in a face-to-back manner, such that two different surface patches are involved in forming an interface (Fig. 1a). Therefore, we expect many more dihedral than cyclic tetramers to share evolutionary relationships with dimers. This is illustrated by the pathway from a dimer to a dihedral tetramer (Fig. 1a) and the disallowed transition from a dimer to a cyclic tetramer.

Following this idea, we looked at evolutionary relationships in terms of sequence similarity between different quaternary structures to unveil the routes most commonly taken to build larger complexes (Fig. 2b). Each quaternary structure is represented schematically with the numbers of proteins of each type. Pairs of quaternary structures are connected according to the statistical significance of the number of evolutionary transitions between them. Most pairs have fewer transitions between them than expected in a random model (Methods) as exemplified by monomers ( $C1$ ) and dihedral tetramers ( $D2$ ). Other pairs with insignificant numbers of transitions are shown in Supplementary Fig. 2. We find that cyclic dimers, trimers and tetramers share notable numbers of transitions with their dihedral counterparts, supporting the stepwise evolutionary scenario where homomers with dihedral symmetry evolve through cyclic intermediates (Fig. 1a).

Notably in this stepwise scenario, two evolutionary routes lead to a dihedral complex ( $D_n$ ): either from  $n$  dimers or from two cyclic  $n$ -mers (Fig. 2b). This raised the question as to whether it was possible to identify which of these two routes was taken by a given dihedral complex. On the basis of energetic considerations (Supplementary Information 1), we propose that a hierarchy of interface sizes exists within dihedral complexes, and that the larger interface is conserved in evolution. To test this hypothesis, we looked for tetramers homologous to a dimer, as well as hexamers homologous to a dimer or trimer. In this data set (Fig. 2c and Supplementary Table 1) we examined whether the interface within the dimer or trimer corresponded to the largest interface in the homologous tetramer or hexamer. Among 33 tetramers and 19 hexamers studied, 49 complexes conserve the larger interface with the dimeric or trimeric homologue, whereas only 3 conserve their smaller interface (Fig. 2c and Supplementary Table 1). This result implies that the evolutionary route of a homomer can be predicted solely from its interface sizes. Our predictions for the evolutionary pathways of  $D3$ ,  $D4$  and  $D5$  complexes (Supplementary Fig. 3a) have led us to formulate a general model of homomer evolution (Supplementary Fig. 3b).

It is notable that this signature of complex formation (hierarchy in interface sizes) is conserved throughout evolution. This can be interpreted in at least two different although not mutually exclusive ways: (1) once the complex is formed there is no need to dramatically change the interface size, analogous to the classical explanation for the marginal stability of proteins<sup>20</sup> (that is, selective pressure becomes almost non-existent beyond the point where proteins fold); and (2) maintaining a hierarchy of interface strengths is important for a precise order during assembly<sup>21,22</sup>, in which case the largest interface would reflect the main intermediate species during assembly. To test this hypothesis we targeted ten complexes for study using electrospray mass spectrometry (Fig. 4a and Supplementary Table 2).

Initially we verified that the complexes could be generated intact and corresponded to the stoichiometry described in the protein data bank (PDB). The mass spectra recorded for two hexamers with  $D3$  symmetry and one 14-mer with  $D7$  symmetry revealed that the intact homomer is maintained in each case (Fig. 4c). We then induced the disassembly of each complex through the careful change in ionic strength or the stepwise addition of partial denaturants. We detected stable subcomplexes corresponding to trimers and dimers for

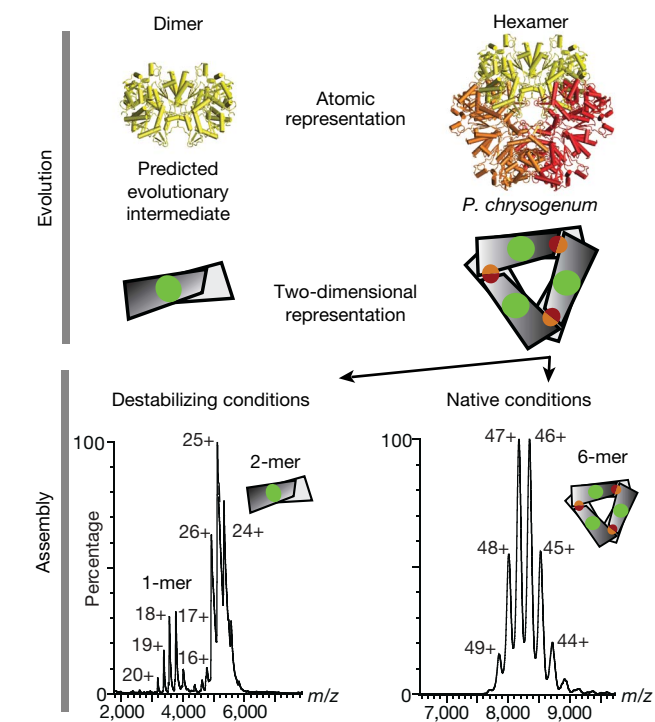
hexameric AUH protein (an RNA binding protein), and MoaC (a molybdenum cofactor biosynthesis protein), respectively (Supplementary Table 2). Examination of the interface size shows that in both cases the larger interface is maintained. Similarly for the  $\text{Ca}^{2+}$ -dependent kinase with D7 symmetry, a dimer is the principal dissociation product and buries the largest interface (Fig. 4c). For one complex (PDB entry 1vea) our results were ambiguous as no intermediate and only monomeric subunits were detected; for another complex (PDB entry 1umg) we predicted a tetramer and detected a dimer. In this case, both subcomplexes bury large surfaces ( $>5,000\text{\AA}^2$ ), which may bias the use of interface size as a proxy for interface strength. For the remaining complexes, the predicted subcomplex containing the larger interface was observed. These results demonstrate that the largest interface is maintained consistently during disassembly.

To address whether the disassembly process was the reverse of the assembly pathway, we attempted to reassemble a subset of the complexes studied by dilution of the denaturant and/or manipulation of the ionic strength. In  $\sim 50\%$  of the complexes examined we were able to reassemble the original homomer. These results—together with previous studies where reassembly was found to be strongly dependent on factors such as ionic strength, temperature and concentration of denaturant<sup>23,24</sup>—indicate that disassembly is the reverse of assembly under the appropriate conditions.

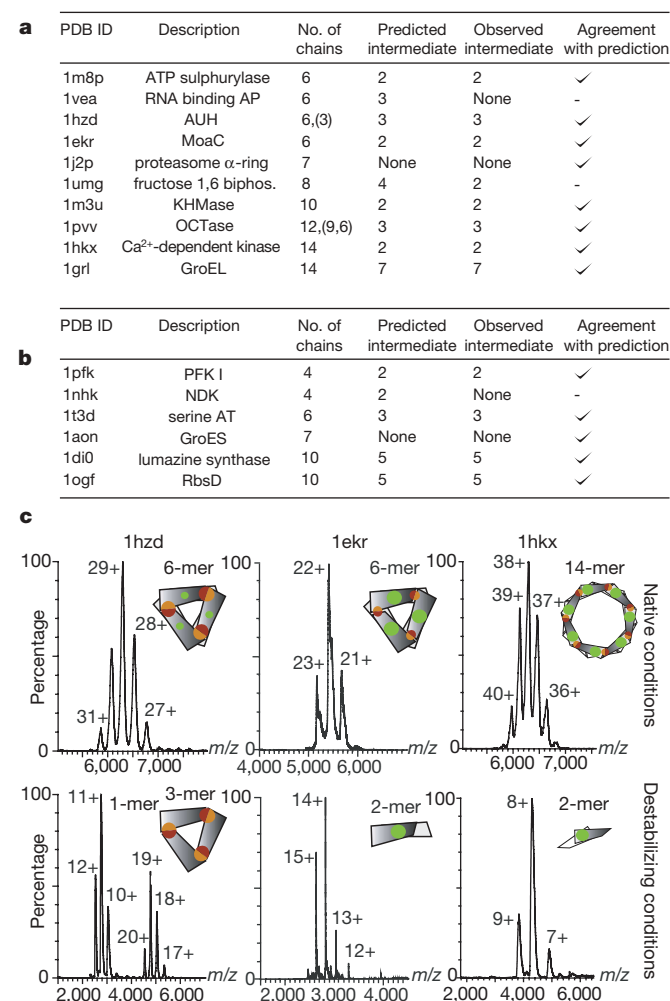
To complement our experimental observations, we found six additional complexes for which (dis)assembly intermediates had been reported (Fig. 4b). Of these, five match our prediction and one (nucleoside diphosphate kinase) had no intermediate detected. This homomer may either assemble without forming subcomplexes, or subcomplexes may have escaped detection. Alternatively,

formation of subcomplexes might involve factors absent from the experimental set-up<sup>25</sup>. Thus, although there are exceptions, we find agreement between the evolutionary pathway and (dis)assembly pathway in 81% of the cases we examined.

Overall, through analysis of a large set of homomers, we have shown that the evolutionary pathway of a homomer can be inferred from its atomic structure morphology. This allowed us to predict the (dis)assembly pathway of homomers in solution, and design mass-spectrometry-based experiments to validate our predictions. Results revealed that the (dis)assembly pathway, which takes place on a protein-folding timescale ( $\sim$ seconds), mimics the evolutionary pathway that has taken place over a considerably longer timescale ( $\sim$ millions of years). This is the first time that a general principle for formation and assembly of homomers has been demonstrated. We hope that this will stimulate further studies, as relationships between



**Figure 3 | Prediction of evolutionary routes and link with (dis)assembly in solution.** ATP sulphurylase is a hexamer in *Penicillium chrysogenum*, with a predicted dimeric evolutionary intermediate based on interface sizes (that is, the interface in the dimer (green patch) is larger than the trimeric interface (red and orange patch; top panel)). We perturb the hexameric ATP sulphurylase (PDB accession 1m8p, Supplementary Table 2) to disassemble it into subcomplexes probed using electrospray mass spectrometry (bottom panel) and determine whether a dimeric (with the larger interface) or a trimeric (small interface) subcomplex is detected. Both dimers and monomers were detected, with no evidence of trimers.



**Figure 4 | (Dis)assembly pathways in 16 complexes.** **a**, Homomers for which (dis)assembly was probed using electrospray mass spectrometry (Methods). ‘None’ denotes no intermediate detected. Complexes agree with our prediction where the subcomplex containing the larger interface is the most stable in eight out of ten cases. **b**, List of homomers for which: (1) information on (dis)assembly has been reported (Supplementary Table 3); (2) a crystal structure is known; and (3) the intermediate species observed during (dis)assembly could be mapped to a subcomplex in the structure. For these complexes, we found an agreement with our prediction in 5/6 cases. **c**, Mass spectra showing intact complexes (top panel) as well as subcomplexes obtained after destabilization in solution (bottom panel). AUH, an RNA binding protein; GroEL and GroES are chaperonins; KHMase, ketopantoate hydroxymethyltransferase; MoaC, molybdenum cofactor biosynthesis protein; RNA binding AP, RNA binding antitermination protein; serine AT, serine acyltransferase; OCTase, ornithine carbamoyltransferase.

folding, complex formation and aggregation are only beginning to be explored.

## METHODS SUMMARY

**Data set of homomers.** All data sets of homomers used were derived from the 3D complex database<sup>8</sup>. As the quaternary structure annotation in the PDB biological unit is erroneous in some cases, we used a manually curated data set<sup>26</sup>.

**Randomization of evolutionary routes.** To assess the significance of the number of evolutionary relationships between proteins with different quaternary structures, we compared the observed numbers to a random model of quaternary structure transitions where evolutionary relationships are reassigned randomly in proportion to the size of each quaternary structure type.

**Prediction of evolutionary routes.** The size of an interface is given by the number of amino acids in contact, as defined previously<sup>8</sup>. We predict evolutionary intermediates by taking the 'closed' subcomplex containing the largest interface. In cyclic complexes with three or more subunits, each subunit buries two equivalent surfaces. Thus, these interfaces are counted twice when compared to dimer interfaces.

**Intact complexes.** Complexes were donated by crystallographers and taken from a random selection from the PDB. For further details see Methods.

**Generating subcomplexes.** Intact complexes were disrupted through change in ionic strength or the stepwise addition of dimethylsulphoxide, methanol or acetonitrile. This process is illustrated in Supplementary Fig. 4 and solution conditions are summarized in Supplementary Table 2.

**Full Methods** and any associated references are available in the online version of the paper at [www.nature.com/nature](http://www.nature.com/nature).

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**Supplementary Information** is linked to the online version of the paper at [www.nature.com/nature](http://www.nature.com/nature).

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**Author Contributions** E.D.L., E.B.E., C.V.R. and S.A.T. designed the experiments and wrote the manuscript; E.D.L. and E.B.E. performed the bioinformatics and mass spectrometry experiments, respectively.

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## METHODS

**Data set of homomers and symmetry information.** The ~5,000 structures data set used throughout the study is non-redundant at 80% sequence identity. The data set was controlled for a possible bias in the distribution of the number of subunits. As no bias was found<sup>8</sup>, we can be confident in the accuracy of the relative abundances of symmetries as well as their evolutionary relationships. However, an important bias in structural data is the under-representation of membrane proteins, which is discussed further in Supplementary Information 2. For the analysis on quaternary structure conservation, we derived several non-redundant sets of protein pairs. To study conservation within the identity range  $X\% - X + 10\%$ , we used a data set non-redundant at  $X + 20\%$  (with the exception of  $X = 90$  and  $100\%$ ). All data sets and the symmetry information were derived from the 3D complex database<sup>8</sup>.

**Randomization of evolutionary routes.** To assess the significance of the number of evolutionary relationships between proteins with different quaternary structures, we devised a random model of quaternary structure transitions. In this model, evolutionary relationships are reassigned randomly in proportion to the size of each quaternary structure type. For each evolutionary link between two quaternary structure types, a first quaternary structure type is picked up with a probability  $p(QS) = T^{QS} / T$ , where  $T^{QS}$  is the quaternary structure size (number of proteins), and  $T$  is the total number of proteins. A second quaternary structure is chosen in the same way but the type picked first is set aside and cannot be selected again. One-hundred rounds of reassignment were performed, and a mean number of links and associated standard deviation were calculated for each quaternary structure pair.

**Prediction of evolutionary routes.** To decompose the complexes into their evolutionary intermediates, we first grouped together interfaces related by a symmetry operation. We then ranked each group according to the average size of interfaces it contained. The size is given by the number of amino acids in contact as defined previously<sup>8</sup>. The complex was broken by removing each group of interfaces one by one, starting with the weakest. After removal of each group, we checked if all the subunits in the complex were still connected via the remaining interfaces. When the complex breaks down, the subcomplexes found correspond to the predicted evolutionary intermediates. Note that in cyclic complexes with three subunits or more, each subunit buries two equivalent surfaces. Thus, these interfaces are counted twice when compared to interfaces within dimers.

**Probing the (dis)assembly pathway using electrospray mass spectrometry.** Methanol and acetonitrile were obtained from Fisher scientific; ammonium acetate and dimethylsulphoxide were from Sigma. All chemicals used were American Chemical Society or HPLC grade and water was obtained from an ELGA LabWater's PURELAB Maxima system.

Before mass spectrometry, complex-containing solutions were desalted and concentrated by centrifugation at  $10,000g$  in Vivaspin concentrator tubes (exclusion limits 5,000, 10,000, 30,000; Vivaspin, Sartorius) to a final concentration of 20–55  $\mu M$  of protein complex (Supplementary Table 2). The intact complex was diluted with ammonium acetate to 3–5  $\mu M$  immediately before the mass spectrometry analysis. Two microlitres of complex-containing solutions were analysed using nanoelectrospray and a quadrupole time-of-flight mass spectrometer (QSTAR, Sciex). The instrument was modified for the detection of high masses<sup>27</sup>. For nanoelectrospray, gold-coated borosilicate capillaries were prepared in-house as described previously<sup>28</sup>. The following instrumental parameters were used: capillary voltage up to 1.5 kV, declustering potential 200 V, focusing potential 250 V, declustering potential-2 15 V and collision energy up to 280 V, microchannel plate detector 2350V. Argon was used as a collision gas for tandem mass spectra. All spectra were calibrated externally using caesium iodide ( $100 \text{ mg ml}^{-1}$ ).

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